



Bioorganic & Medicinal Chemistry Letters 16 (2006) 4917-4921

Bioorganic & Medicinal Chemistry Letters

Tetrahydroisoquinolines as MCH-R1 antagonists

T. K. Sasikumar,* L. Qiang, W.-L. Wu, D. A. Burnett, W. J. Greenlee, K. O'Neill, B. E. Hawes, M. van Heek and M. Graziano

Schering-Plough Research Institute, 2015 Galloping Hill Road, Kenilworth, NJ 07033, USA

Received 16 May 2006; revised 14 June 2006; accepted 14 June 2006

Available online 7 July 2006

Abstract—A series of potent and selective inhibitors of h-MCH-R1 has been developed based on the piperidine glycineamide compounds I and II. These structurally more rigid tetrahydroisoquinolines (III and IV) showed better pharmacokinetics. The highly potent compounds 12d and 12g displayed excellent rat pk. © 2006 Elsevier Ltd. All rights reserved.

Melanin concentrating hormone (MCH) is a 19-membered neuropeptide that is found in the lateral hypothalamus and regulates food intake. 1,2 There is evidence for involvement of MCH in feeding and obesity.3 One of the major findings is that hypothalamic MCH peptide levels increase during fasting in ob/ob and WT mice. ICV administration of MCH or analogs stimulates feeding in rodents and MCH-/- mice are hypophagic and leaner than WT mice but otherwise healthy. 4 MCH receptor knock-out mice are lean, hypophagic, hyperactive, have reduced fat mass, have increased metabolic rate, and they are resistant to diet-induced obesity (DIO). Evidence from knock-outs suggests an MCH receptor antagonist should be beneficial for treatment of obesity and related disorders.^{5,6} Several classes of small molecule MCH-R1 antagonists have recently been disclosed. 7-12

Recently we found compounds of the types I and II are potent and selective MCH-R1 antagonists useful for the treatment of metabolic diseases. Compounds of this piperidine glycineamide series had been hindered by moderate pharmacological properties primarily due to the amide hydrolysis. In order to minimize these issues, we designed constrained analogs III and IV as shown in Figure 1. This restriction would better define the active binding conformations and mask the glycineamide structure. Since the free basic N–H is tied back to the aromatic ring, we anticipated less metabolism among these tetrahydroisoquinoline (THQ) structures (III and IV).

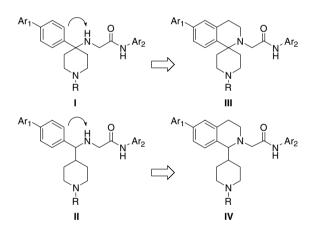


Figure 1. Design of tetrahydroisoquinoline MCH antagonists III and ${\bf IV}$.

The synthesis of various spirocyclic as well as 2-substituted tetrahydroisoquinoline structures and structure—activity relationships (SARs) are described in this paper.

The spirocyclic tetrahydroisoquinoline compound **2** was synthesized from 3-methoxyphenethylamine **1** by Pictet–Spengler cyclization. ¹⁴ During this reaction, the *tert*-but-oxy carbonyl group was hydrolyzed. The methyl group was removed by the reaction of BBr₃ and the Boc group was re-introduced in good yield. The phenol was converted to the triflate and Suzuki coupling reaction on this intermediate afforded the biaryl compound **3**. ¹⁵ N-Alkylation using methylbromoacetate gave compound **4**. Sodium hydride or trimethylaluminum mediated displacement of methyl ester with aromatic anilines afforded the corresponding amides in moderate

Keywords: Melanin concentrating hormone; Tetrahydroisoquinolines. *Corresponding author. Tel.: +1 908 740 4373; fax: +1 908 740 7164; e-mail: thavalakulamgar.sasikumar@spcorp.com

yields. ¹⁶ The Boc deprotection was achieved by trifluoroacetic acid and the reductive alkylation using a wide variety of aldehydes and ketones under standard reaction conditions furnished the final target compounds 5a-q in good yields (Scheme 1).

The MCH-R1 affinities of several representative spirocyclic glycineamide compounds containing modifications on the piperidine nitrogen are shown in Table 1. A wide range of alkyl substitutions on the piperidine nitrogen is tolerated. The cyclopropylmethyl 5f, cyclopentyl 5g, cyclobutyl 5i, and cycloheptyl 5j were the best among the several other compounds prepared. Acylations and sulfonylations of the piperidine nitrogen completely eliminate the MCH-R1 binding affinity (5n-q).

Scheme 1. Reagents and conditions: (a) *N*-Boc-piperidone, H₃PO₄, 90 °C; (b) BBr₃, CH₂Cl₂; (c) Boc₂O; (d) PhNTf₂, CH₂Cl₂; (e) 3-CN-phenylboronic acid, Pd(PPh₃)₄, Na₂CO₃, Tol/MeOH, 90 °C; (f) BrCH₂COOMe, K₂CO₃, CH₃CN; (g) 3,5-di-Cl-aniline, NaH, THF; (h) TFA, CH₂Cl₂; (i) RCHO, NaBH(OAc)₃, CH₂Cl₂.

Table 1. MCH-R1 binding affinities of spirocyclic THQs (5a-q)

Compound	R	h-MCH-R1 $K_{\rm i}^{\rm a}$ (nM)	
5a	Boc	>1000	
5b	Н	59	
5c	Methyl	38	
5d	Ethyl	38	
5e	Isopropyl	29	
5f	Cyclopropylmethyl	15	
5g	Cyclopentyl	16	
5h	1-Hydroxyethyl	41	
5i	Cyclobutyl	13	
5j	Cycloheptyl	11	
5k	1-Tetrahydro-3-thienyl	34	
51	1-Tetrahydropyran-4-yl	16	
5m	3-Furanylmethyl	36	
5n	Acetyl	823	
50	Methylsulfonyl	>1000	
5p	1-Dimethylaminosulfonyl	>1000	
5q	1-Ethylaminosulfonyl	>1000	

^a Values are means of three experiments. Variability around the mean value was <5%.

Next, we turned our attention to the alterations of the aromatic amide group in order to optimize the right-hand side of the molecule. Deprotection of **4** followed by reductive alkylation and subsequent amidation using various anilines furnished compounds **6a–j** (Scheme 2). As seen from Table 2, 3,5-dichlorophenyl glycineamide compound **5f** still has the best MCH-R1 binding affinity ($K_i = 15 \text{ nM}$). Other aromatic amides such as 3-Cl-4-F-phenyl **6d** and 3-CF₃-4-F-phenyl **6h** also showed a similar binding profile. We decided to keep the 3,5-dichlorophenyl group as a constant in the further development of SAR in the THQ series.

After having examined the binding affinity of spirocyclic compounds, we began looking into the homologated tetrahydroisoquinoline structures represented by **IV** (Fig. 1). Compound of this type was prepared according to Scheme 3. At first we decided to study the biaryl SAR in detail. Pictet—Spengler cyclization of 1 with *N*-cyclopentylpiperidine-4-carboxaldehyde afforded compound 7. Initial trials of this cyclization reaction using phosphoric acid gave mixture of products. However, cyclization reaction in boiling TFA gave 7 in 60% yield. Further modifications of 7 according to Scheme 3 afforded compounds **10a**–**p** (Table 3).

Scheme 2. Reagents and conditions: (a) TFA, CH₂Cl₂; (b) RCHO, NaBH(OAc)₃, CH₂Cl₂; (c) Ar-NH₂, NaH, THF.

Table 2. MCH-R1 binding affinities of spirocyclic THQs (6a-i)

Compound	Ar	h-MCH-R1 K_i^a (nM)
5f	3,5-Dichlorophenyl	15
6a	3,5-Difluorophenyl	63
6b	3,4-Dichlorophenyl	138
6c	3-CF ₃ -4-Cl-phenyl	119
6d	3-Cl-4-F-phenyl	24
6e	4-Cl-phenyl	138
6f	3-Cl-phenyl	45
6g	3,4-Difluorophenyl	35
6h	3-CF ₃ -4-F-phenyl	25
6i	3-CF ₃ -5-F-phenyl	54

^a Values are means of three experiments. Variability around the mean value was <5%.

Scheme 3. Reagents and conditions: (a) *N*-cyclopentylpiperidine-4-carboxaldehyde, TFA, reflux; (b) BrCH₂CONHAr, K₂CO₃, CH₃CN; (c) BBr₃, CH₂Cl₂; (d) PhNTf₂, CH₂Cl₂; (e) ArB(OH)₂, Pd(PPh₃)₄, Na₂CO₃, Tol/MeOH.

Table 3. MCH-R1 binding affinities of THQs (10a-p)

Compound	Ar	h-MCH-R1 K_i^a (nM)
10a	3-CN-phenyl	11
10b	Phenyl	168
10c	4-CN-phenyl	108
10d	3-F-phenyl	73
10e	3-Cl-phenyl	43
10f	3-MeO-phenyl	95
10g	3-CF ₃ -phenyl	191
10h	3-CF ₃ O-phenyl	77
10i	3-CHO-phenyl	77
10j	3,6-Di-Cl-phenyl	87
10k	8-Quinolinyl	799
10l	4-Pyridyl	24
10m	3-Pyridyl	23
10n	3-(1 <i>H</i> -Imidazol-2-yl)phenyl	46
10o	1 <i>H</i> -Pyrrol-2-yl	455
10p	3-Thienyl	115

 $^{^{\}rm a}$ Values are means of three experiments. Variability around the mean value was $<\!5\%$.

Scheme 4. Reagents and conditions: (a) *N*-benzylpiperidine-4-carbox-aldehyde, TFA, reflux; (b) BrCH₂CONHAr, K₂CO₃, CH₃CN; (c) BBr₃, CH₂Cl₂; (d) PhNTf₂, CH₂Cl₂; (e) ArB(OH)₂, Pd(PPh₃)₄, Na₂CO₃, Tol/MeOH; (f) chloroethylchloroformate, CH₂Cl₂; (g) RCHO, NaBH(OAc)₃, CH₂Cl₂.

Table 4. MCH-R1 binding affinities of THQs (12a-q)

Compound	R	h-MCH-R1 K_i^a (nM)	
12a	Н	26	
12b	Methyl	22	
12c	Phenylmethyl	24	
12d	5-(OH)-1-pentyl	6.8	
12e	Cycloheptyl	9.7	
12f	N-Methylpiperidinyl	68	
12g	1-Tetrahydropyran-4-yl	6.4	
12h	1-Tetrahydro-3-thienyl	7.0	
12i	3-Furanylmethyl	14	
12j	2-MeO-phenylmethyl	16	
12k	Cyclopropylmethyl	9.3	
121	Cyclobutyl	15	
12m	Phenylpropyl	69	
12n	Isopropyl	35	
12o	Cyclohexyl	10	
12p	Acetyl	927	
12q	Methanesulfonyl	718	

^a Values are means of three experiments. Variability around the mean value was <5%.</p>

As evidenced from Table 3, the biaryl portion of the molecule is less tolerant of substitutions. 3-Cyanophenyl moiety is still the best substitution in the biaryl region of the molecule. Groups like 3- and 4-pyridyls are tolerated to some extent (10l and 10m). Heterocyclic substitutions such as 2-pyrrolyl 10o and quinolyl 10k reduce the binding affinity by 40- to 70-fold. 3-Chlorophenyl substitution 10e afforded a 4-fold decrease in potency relative to 3-cyanophenyl.

The piperidine nitrogen SAR was studied according to Scheme 4. Pictet-Spengler cyclization of 1 with

13, MCH-R1
$$K_i$$
 = 133 nM 14, MCH-R1 K_i = 84 nM

Figure 2. Benzimidazole compounds 13 and 14.

Table 5. PK data for selected compounds

Compound	h-MCH-R1 K _i ^a (nM)	Rat PK (10 mpk, po) ^b AUC (ng h/mL)
5g	16	332
10a	11	910
12d	6.8	1636
12g	6.4	1244
12h	7	499

^a Values are means of three experiments. Variability around the mean value was <5%.

N-benzylpiperidine-4-carboxaldehyde afforded compound 11. N-Alkylation of 11 followed by demethylation, triflation, and Suzuki coupling produced the biaryl compound, and subsequent debenzylation and reductive alkylations using various aldehydes and ketones gave compounds 12a–q (Scheme 4).

The SAR of the MCH-R1 binding of several tetrahydro-isoquinoline structures with substitutions on the piperidine nitrogen is summarized in Table 4. Generally reductive alkylation products showed excellent MCH-R1 binding affinity. Compounds such as hydroxypentyl 12d (K_i 6.8 nM), tetrahydropyranyl 12g (K_i = 6.4 nM), tetrahydrothienyl 12h (K_i = 7.0 nM), and cyclopropyl methyl 12k (K_i = 9.3 nM) are some of the very active compounds in this series. Other modifications to the piperidine nitrogen, including acylation, sulfonylation, and urea formation, resulted in inactive compounds. These results indicate that the basic nitrogens both in chemotypes III and IV are very important for MCH-R1 binding affinity. In most of the cases, the MCH-R1 SAR for chemotypes IV parallels that for chemotype III.

We briefly attempted to change the amide portion of the THQ core by introducing amide isosteres such as benzimidazoles and oxazoles. These attempts did not provide any superior MCH-R1 compounds compared to the amide derivatives. Some examples are given in Figure 2. Benzimidazole derivatives 13 and 14 showed moderate MCH-R1 binding affinity (Fig. 2).

Compounds with good MCH-R1 affinity were selected for PK studies (Table 5). Compounds like **12d** and **12g** showed remarkable improvement in pharmacokinetics. The corresponding uncyclized piperidine glycineamides showed zero or negligible rat AUC under similar experimental conditions.¹⁷ These results indicate that con-

formationally restricted analogs are better MCH-R1 antagonists with improved pharmacokinetic profiles. Further in vivo results will be reported in due course.

In summary, we have undertaken a three-point modification of our tetrahydroisoquinoline (THQ) core structures and generated a number of selective MCH-R1 antagonists. Both spirocyclic as well as homologated tetrahydroisquinolines followed a similar SAR trend. We have shown that the basic nitrogen of the piperidine moiety is very important for high affinity binding. Pharmacokinetic studies confirmed an enhanced profile relative to non-THQ analogs.

Acknowledgment

We thank Dr. John Clader for helpful discussions.

References and notes

- 1. Qu, D.; Ludwig, D. S.; Gammeltoft, S.; Piper, M.; Pelleymounter, M. A.; Cullen, M. J.; Mathes, W. F.; Przypek, R.; Kanarek, R.; Maratos-Flier *Nature* **1996**, *380*, 243.
- Saito, Y.; Nothacker, H. P.; Civelli, O. Trends Endocrinol. Metab. 2000, 11, 299.
- Schwartz, M. W.; Woods, S. C.; Porte, D., Jr.; Selley, R. J.; Baskin, D. G. *Nature* 2000, 404, 661.
- 4. Della-Zuanna, O.; Presse, F.; Ortola, C.; Duhault, J.; Nahon, J. L.; Levens, N. *Int. J. Obesity* **2002**, *26*, 1289.
- Takekawa, S.; Asami, A.; Ishihara, Y.; Terauchi, J.; Kato, K.; Shimomura, Y.; Mori, M.; Murakoshi, H.; Kato, K.; Suzuki, N.; Nishimura, O.; Fujino, M. Eur. J. Pharmacol. 2002, 438, 129.
- Borowsky, B.; Durkin, M. M.; Ogozalek, K.; Marzabadi, M. R.; DeLeon, J.; Lagu, B.; Heurich, R.; Lichtblau, H.; Shaposhnik, Z.; Daniewska, I.; Blackburn, T. P.; Branchek, T. A.; Gerald, C.; Vaysse, P. J.; Forray, C. Nat. Med. 2002, 8, 779.
- 7. Browning, A. Expert Opin. Ther. Patents 2004, 14, 313.
- 8. Soures, A. J.; Wodka, D.; Gao, J.; Lewis, J. C.; Vasudevan, A.; Gentles, R.; Brodjian, S.; Dayton, B.; Ogiela, C. A.; Fry, D.; Hernandez, L. E.; Marsh, K. C.; Collins, C. A.; Kym, P. R. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 4873.
- McBriar, M. D.; Guzik, H.; Xu, R.; Paruchova, J.; Li, S.; Palani, A.; Clader, J. W.; Greenlee, W. J.; Hawes, B. E.; Kowalski, T. J.; O'Neill, K.; Spar, B.; Weig, B. *J. Med. Chem.* 2005, 48, 2274.
- Palani, A.; Shapiro, S.; McBriar, M. D.; Clader, J. W.; Greenlee, W. J.; O'Neill, K.; Hawes, B. Bioorg. Med. Chem. Lett. 2005, 15, 5234.

^b See Ref. 17 for procedure.

- 11. Palani, A.; Shapiro, S.; McBriar, M. D.; Clader, J. W.; Greenlee, W. J.; Spar, B.; Kowalski, T. J.; Farley, C.; Cook, J.; van Heek, M.; Weig, B.; O'Neill, K.; Graziano, M.; Hawes, B. *J. Med. Chem.* **2005**, *48*, 4746.
- McBriar, M. D.; Kowalski, T. J. Ann. Rep. Med. Chem. 2005, 40, 119.
- 13. Wu, W.-L.; Burnett, D. A.; Spring, R.; Qiang, L.; Sasikumar, T. K.; Domalski, M. S.; Greenlee, W. J.; O'Neill, K.; Hawes, B. E. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 3668.
- 14. Pictet, A.; Spengler, T. Chem. Ber. 1911, 44, 2030.
- 15. For an overview of Suzuki coupling reactions, see Suzuki, A.; Brown, H. C. *Organic Synthesis via Boranes* Aldrich, **2003**; Vol. 3.
- Baiocchi, L.; Picconi, G. Tetrahedron: Assymetry 1991, 2, 231.
- 17. Cox, K. A.; Dunn-Meynell, K.; Korfmacher, W. A.; Broske, L.; Nomeir, A. A.; Lin, C. C.; Cayen, M. N.; Barr, W. H. *Drug Discovery Today* **1999**, *4*, 232.